

# **High prevalence of *Clostridium difficile* in the Western Australian environment**

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# **ÆTIOLOGY OF ANTIMICROBIAL-AGENT-ASSOCIATED COLITIS**

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**THE LANCET, APRIL 15, 1978**



# **Identification of *Clostridium difficile* as a cause of pseudomembranous colitis**

(Accepted 16 February 1978)

**BRITISH MEDICAL JOURNAL**

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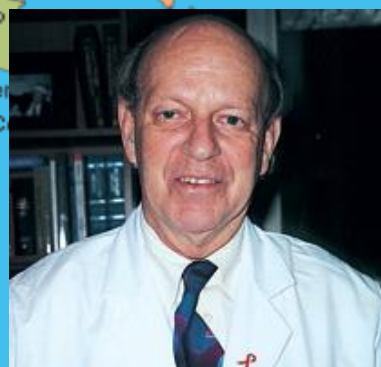
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# **CLOSTRIDIUM DIFFICILE AND THE ÆTIOLOGY OF PSEUDOMEMBRANOUS COLITIS**

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**THE LANCET, MAY 20, 1978**

# **Clindamycin-Induced Enterocolitis in Hamsters as a Model of Pseudomembranous Colitis in Patients**

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# Urgent Threats

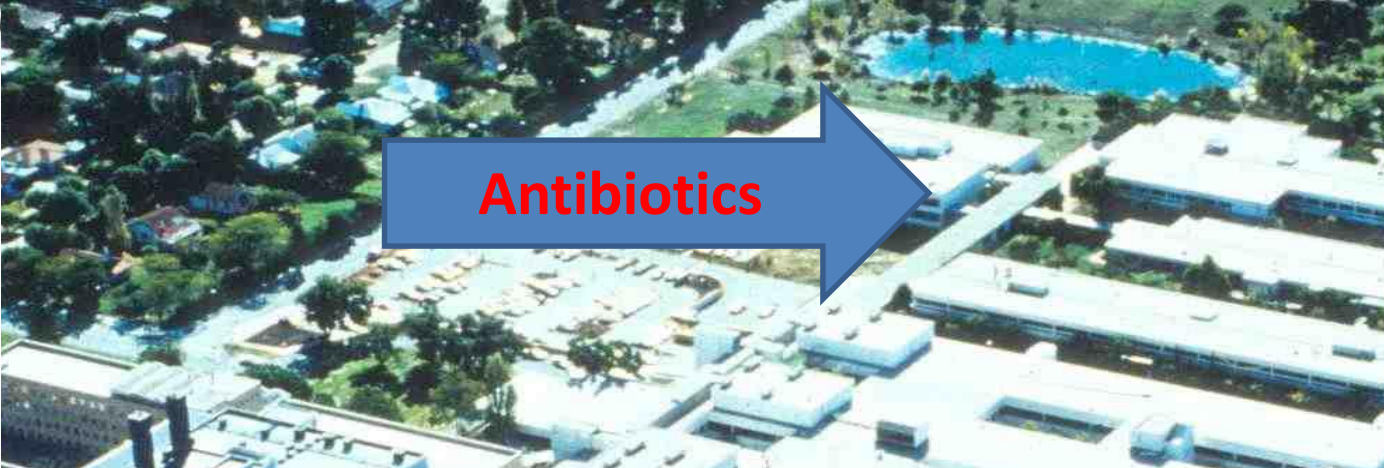
CDC, 2013

- *Clostridium difficile*
- Carbapenem-resistant Enterobacteriaceae (CRE)
- Drug-resistant *Neisseria gonorrhoeae*

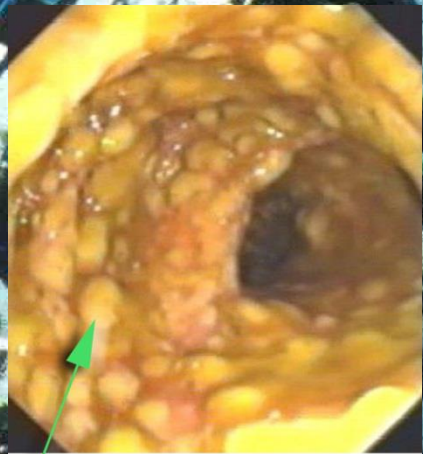


*Clostridium difficile* (*C. difficile*) causes life-threatening diarrhea. These infections mostly occur in people who have had both recent medical care and antibiotics. Often, *C. difficile* infections occur in hospitalized or recently hospitalized patients.





Antibiotics





# The Epidemiology of Community-Acquired *Clostridium difficile* Infection: A Population-Based Study

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## Study highlights

### What is current knowledge?

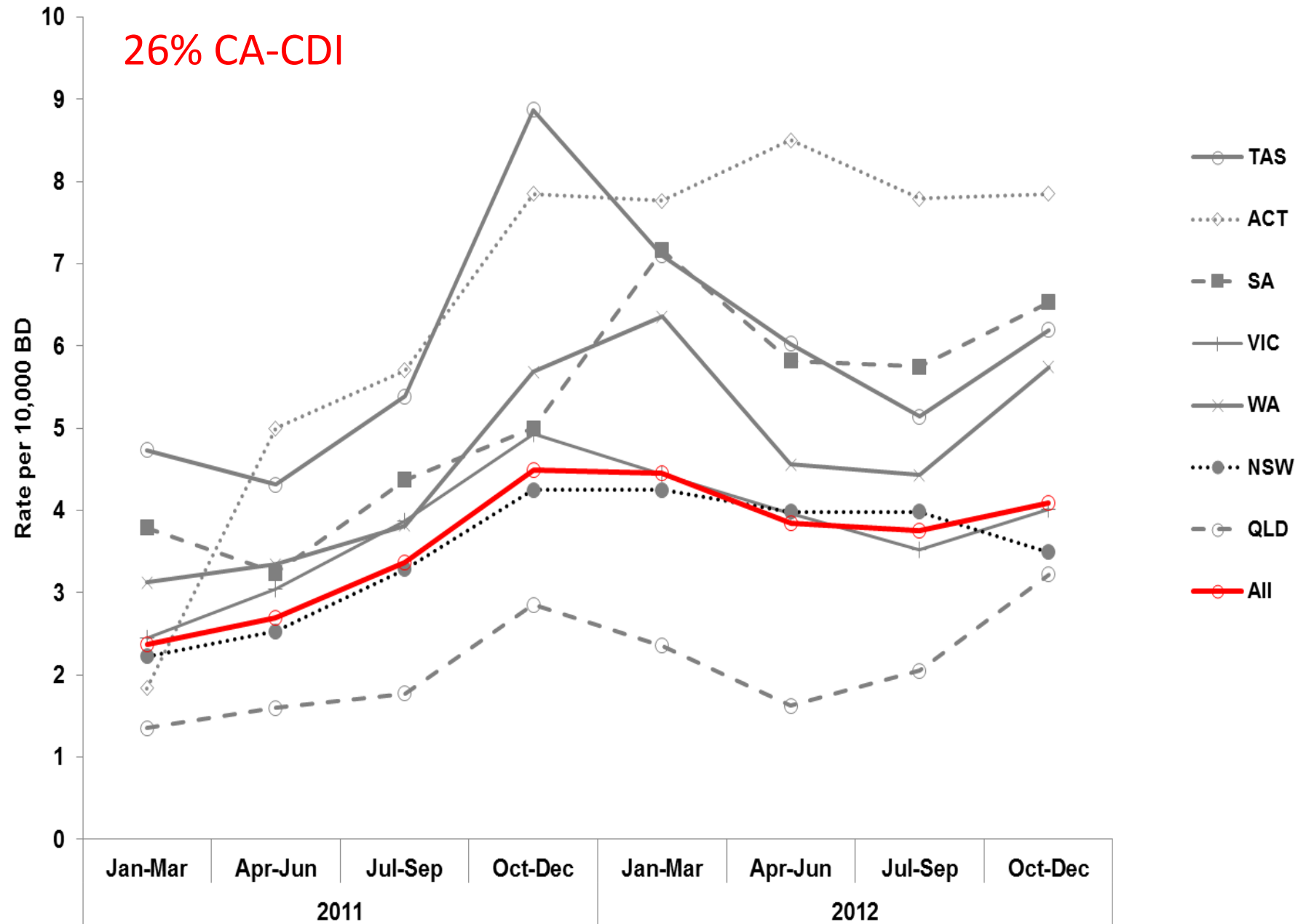
- *Clostridium difficile* infection is increasing worldwide with hospitalization and antibiotic exposure as the most common risk factors.
- The epidemiology and characteristics of community-acquired *Clostridium difficile* infection are not well defined.

### What is new here?

- A major proportion of *Clostridium difficile* infection patients is community-acquired.
- These patients are younger, often lack traditional risk factors, and have less severe disease than patients with hospital-acquired infection.

# Hospital identified CDI in Australia, 2011-2012

(Slimings et al. *Med J Aust* 2014; 200: 272-6)



# *The* NEW ENGLAND JOURNAL *of* MEDICINE

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## Diverse Sources of *C. difficile* Infection Identified on Whole-Genome Sequencing

David W. Eyre, B.M., B.Ch., Madeleine L. Cule, Ph.D., Daniel J. Wilson, D.Phil., David Griffiths, B.Sc., Alison Vaughan, B.Sc., Lily O'Connor, B.Sc., Camilla L.C. Ip, Ph.D., Tanya Golubchik, Ph.D., Elizabeth M. Batty, Ph.D., John M. Finney, B.Sc., David H. Wyllie, Ph.D., Xavier Didelot, D.Phil., Paolo Piazza, Ph.D., Rory Bowden, Ph.D., Kate E. Dingle, Ph.D., Rosalind M. Harding, Ph.D., Derrick W. Crook, M.B., B.Ch., Mark H. Wilcox, M.D., Tim E.A. Peto, D.Phil., and A. Sarah Walker, Ph.D.

### CONCLUSIONS

Over a 3-year period, 45% of *C. difficile* cases in Oxfordshire were genetically distinct from all previous cases. Genetically diverse sources, in addition to symptomatic patients, play a major part in *C. difficile* transmission. (Funded by the U.K. Clinical Research Collaboration Translational Infection Research Initiative and others.)

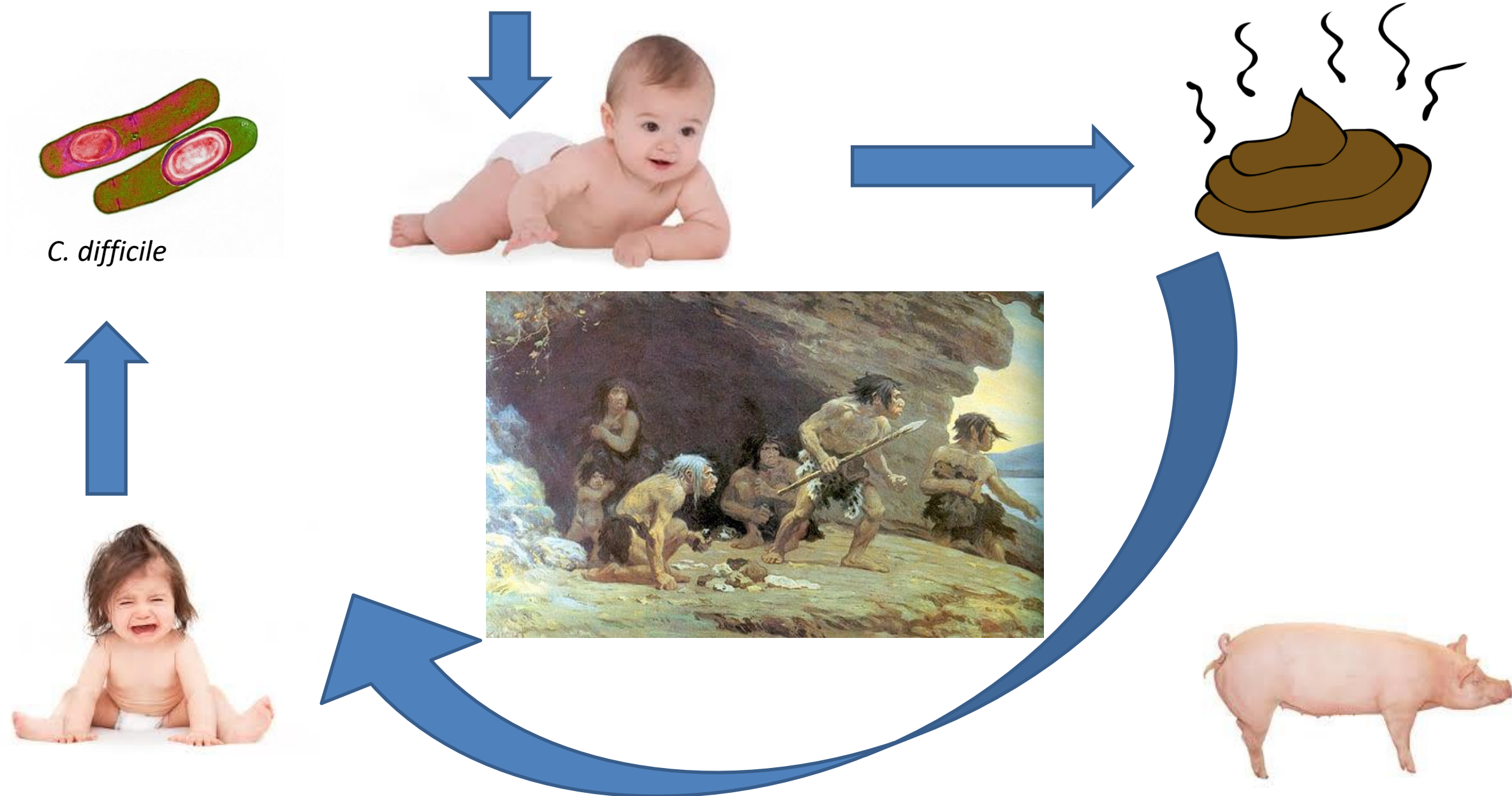
# Risk factors for CDI ?

- Exposure to the organism
- Anything that alters normal gut microflora: exposure to antibiotics – clindamycin, then cephalosporins, some fluoroquinolones
- Others - IBD





# The natural history of *C. difficile*



# DanMap 2007

“This change in prescription habits suggests that the consumption of cephalosporins in pigs is changing from occasional prescription to more systematic prescription in herds producing 14-29% of the weaned pigs.”

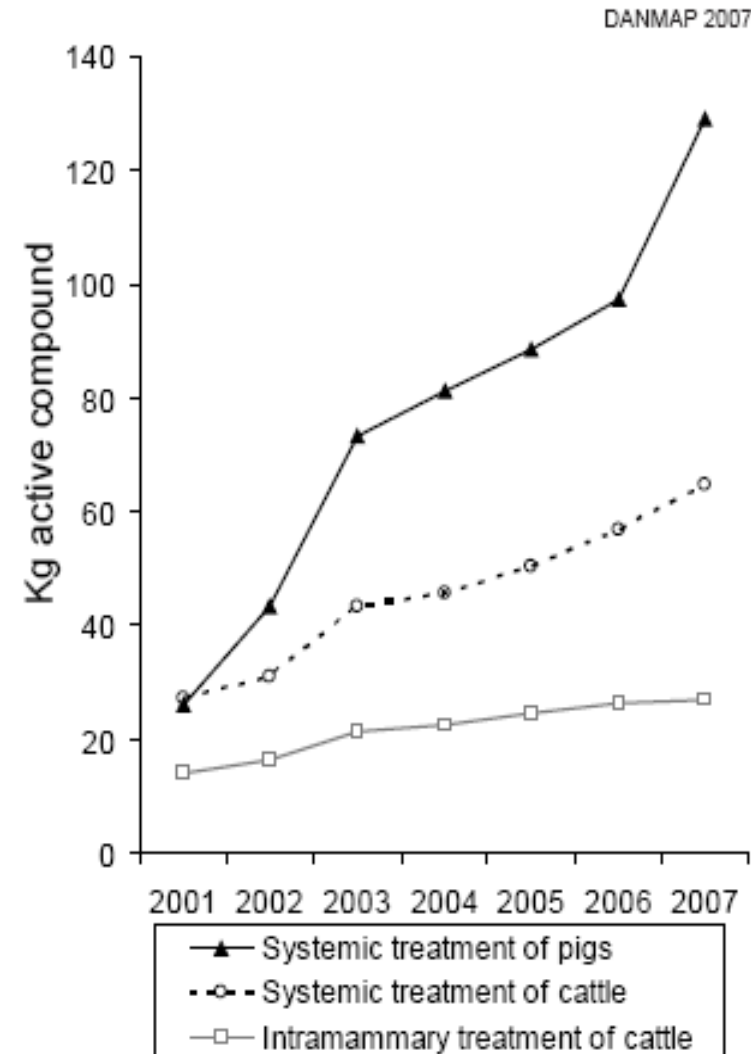


Figure 7. Use of 3rd and 4th generation cephalosporins in pigs and cattle, 2001-2007, Denmark



# Antimicrobial use in the Australian pig industry: results of a national survey

D Jordan,<sup>a\*</sup> JJ-C Chin<sup>a</sup>, VA Fahy,<sup>b</sup> MD Barton,<sup>c</sup> MG Smith<sup>d</sup> and DJ Trott<sup>e</sup>

**Table 2.** Percent of pig herds (n = 197) reported as having used particular antimicrobials in the 12 months prior to survey and the route of administration of those drugs

Antimicrobial	Importance rating for human health	Percent of herds within each category of use				
		Unanswered	Not used	Injection/oral	Oral only	Injection only
Apramycin/neomycin	Medium/low	1.0	47.3	2.5	42.1	7.1
Florfenicol (for respiratory disease)	Low	1.0	95.4	0.5	0.0	3.1
Florfenicol (for gut disease)	Low	2.5	93.4	0.0	0.5	3.6
Lincomycin and spectinomycin	Medium	2.0	64.5	6.1	19.3	8.1
Macrolide	Low	2.0	25.4	32.0	27.9	12.7
Penicillins <sup>a</sup>	Low	1.0	3.6	22.8	4.6	68.0
Ceftiofur	High	1.5	73.7	2.0	0.5	22.3
Tiamulin	Not yet rated	1.0	88.4	1.0	7.1	2.5
Olaquinox	Low	1.0	46.7	0.0	52.3	0.0
Virginiamycin	High	1.5	98.5	0.0	0.0	0.0
Dimetridazole	Medium	2.0	85.3	0.0	12.7	0.0
Sulfonamides	Low	2.0	28.9	5.6	10.7	52.8
Tetracyclines	Low	1.0	15.2	50.8	10.7	22.3
Other antimicrobials		3.1	86.2	0.0	7.6	3.1

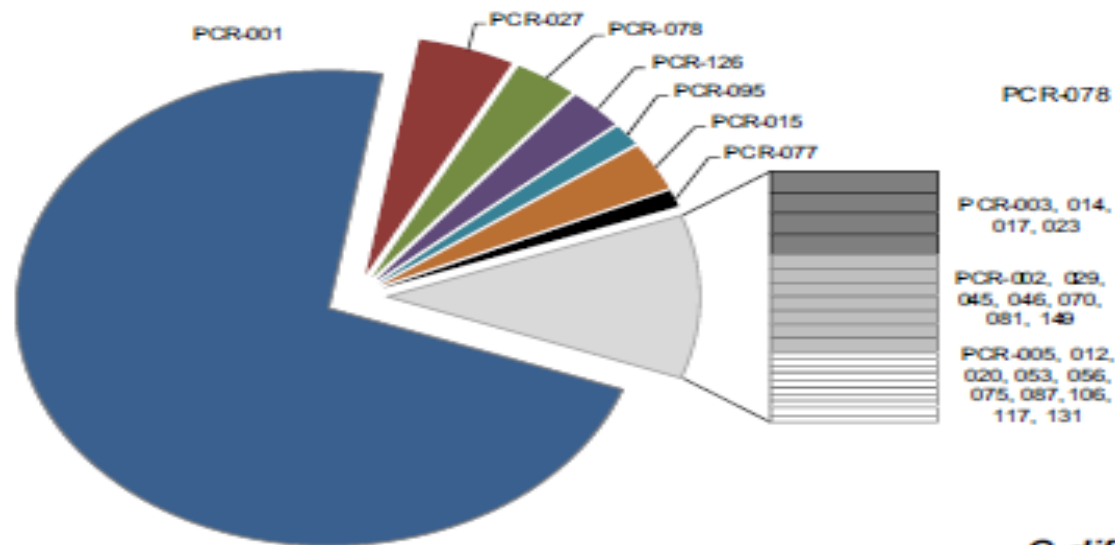
<sup>a</sup>Includes amoxicillin and ampicillin.

Covered about half the large herds in Australia in 2006.

## *C. difficile* in people

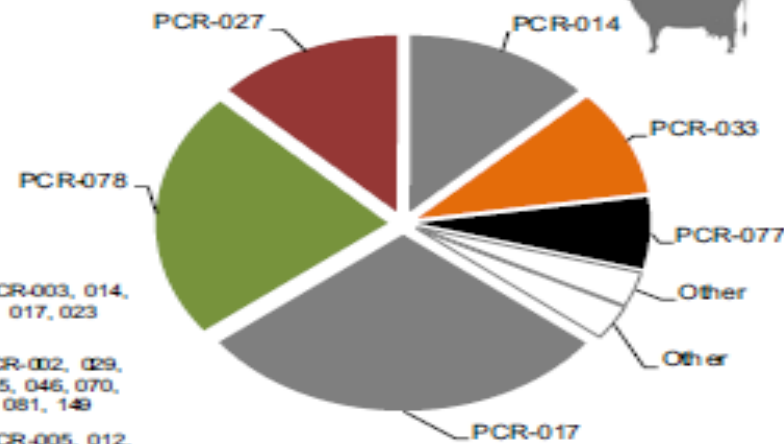


### Hospitals, Germany



## *C. difficile* in calves at the farm, 102 farms, Canada

May-September 2004



## *C. difficile* in foods North America

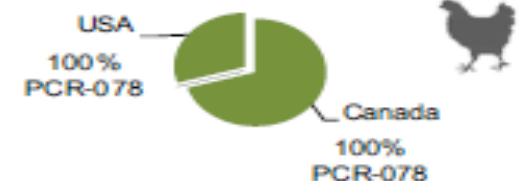
### BEEF meats



### PORK meats



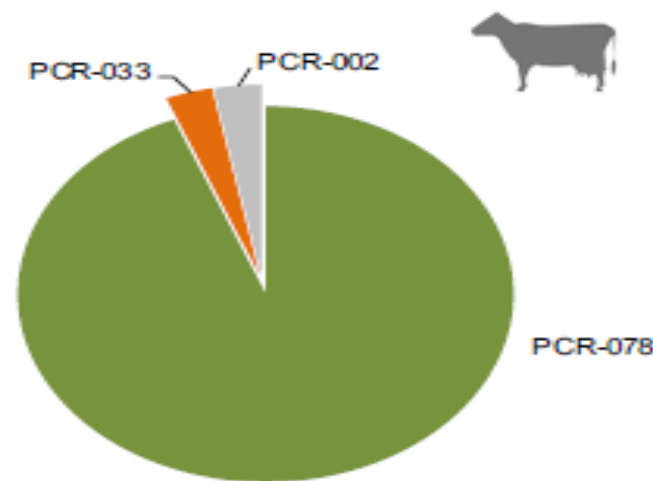
### POULTRY meats



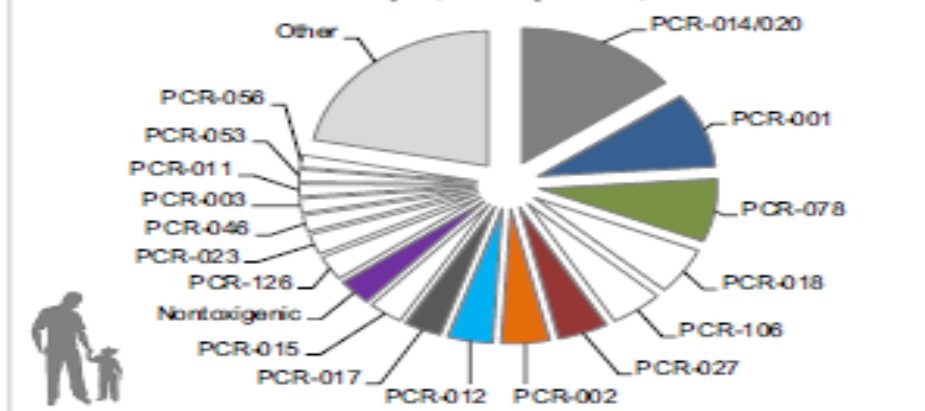
### Seafood & vegetables



## *C. difficile* in calves shipped to a single farm, USA



## *C. difficile* in Europe, hospitals, 34 countries







# Animal strains in Australia

- Ribotype 127 60%
- Ribotype 126 16%
- Ribotype 033 13%



5-7days old

- Ribotype 014 23%
- Ribotype 033 13%
- Ribotype QX009 12%
- Ribotype 237 10%



- Many new ribotypes from animals – CDT+

# Contamination of Australian newborn calf carcasses at slaughter with *Clostridium difficile*

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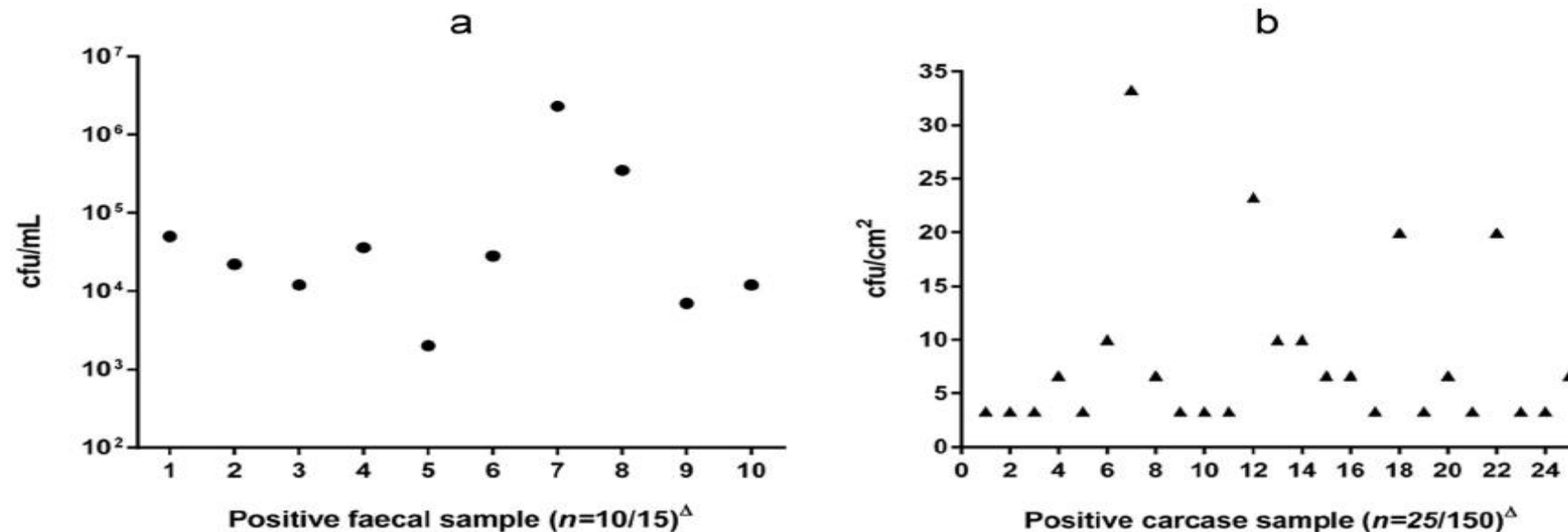


FIG. 1. Concentration of viable *Clostridium difficile* in faeces (a) and on carcass sponges (b). <sup>Δ</sup>Number of isolates above the limit of detection.





# Vegetables in WA


Su-Chen Lim



Sample type	Prevalence
Carrots	1.8-5.3% (1/19)
Onions	1.9-5.6% (1/18)
Beetroots	7.4-22.2% (4/18)
Potatoes	16.6-50.0% (8/16)
Total	6.6-19.7% (14/71)

## High prevalence of *Clostridium difficile* on retail root vegetables, Western Australia

Journal of Applied Microbiology 124, 585-590 © 2017

S.C. Lim<sup>1</sup> , N.F. Foster<sup>2</sup>, B. Elliott<sup>3</sup> and T.V. Riley<sup>1,2,3,4</sup>

PCR ribotype	Toxin gene profile			n (%)
	tcdA	tcdB	cdtA/cdtB	
QX 145	-	-	-	39 (13.7)
<b>UK 101</b>	+	+	-	32 (11.2)
QX 104	-	-	-	30 (10.5)
<b>UK 014/020</b>	+	+	-	18 (6.3)
QX 393	-	-	-	18 (6.3)
QX 142	-	-	-	18 (6.3)
<b>UK 056</b>	+	+	-	17 (6.0)
Novel 3	+	+	-	12 (4.2)
Novel 2	-	-	-	10 (3.5)
<b>UK 012</b>	+	+	-	10 (3.5)
UK 010	-	-	-	10 (3.5)
QX 072	-	-	-	10 (3.5)
UK 051	-	-	-	10 (3.5)
QX 518	-	-	-	10 (3.5)
QX 519	+	+	-	10 (3.5)
<b>UK 002</b>	+	+	-	10 (3.5)
Novel 4	-	-	-	7 (2.5)
<b>UK 237</b>	-	+	+	4 (1.4)
Novel 1	-	-	-	4 (1.4)
<b>UK 137</b>	+	+	-	3 (1.1)
<b>QX 274</b>	+	+	+	2 (0.7)
<b>UK 033</b>	-	-	+	1 (0.4)
Total				285

# Gardening centres

~30% of samples positive for *C. difficile*

Some obvious like animal manures

Some less obvious like  
compost/mulch

But expired vegetables from large  
stores going into compost/mulch



Su-Chen Lim

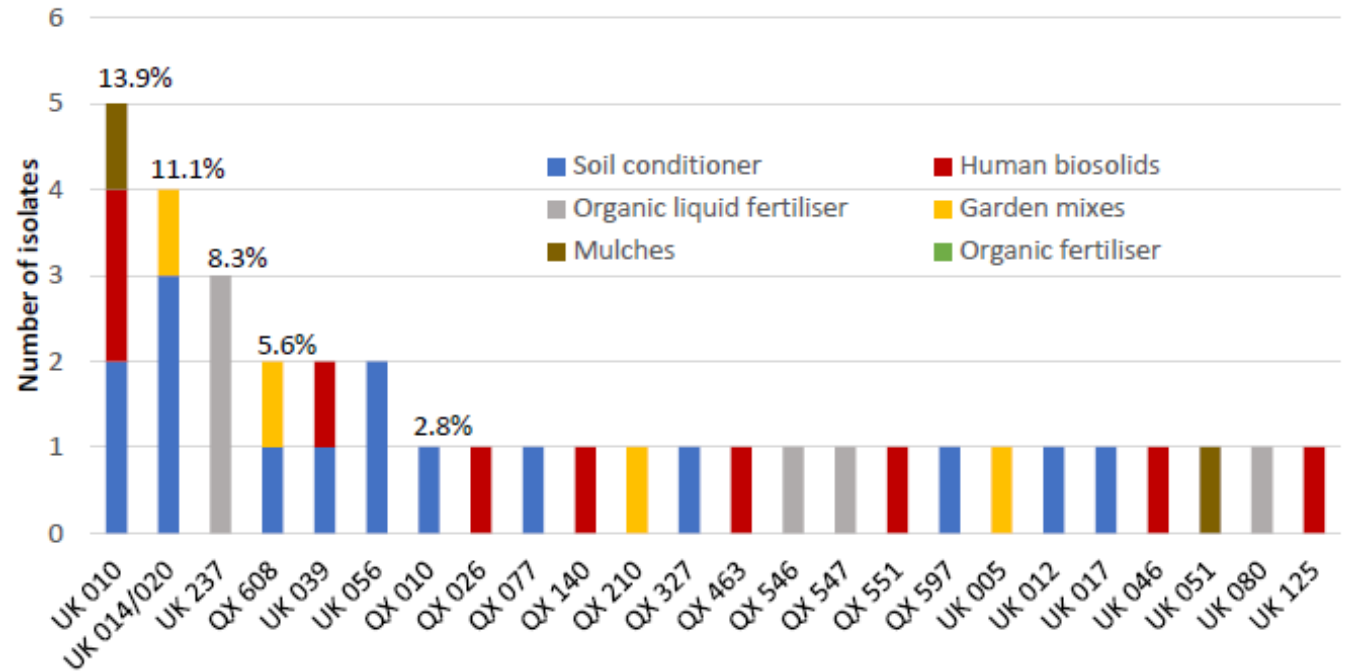
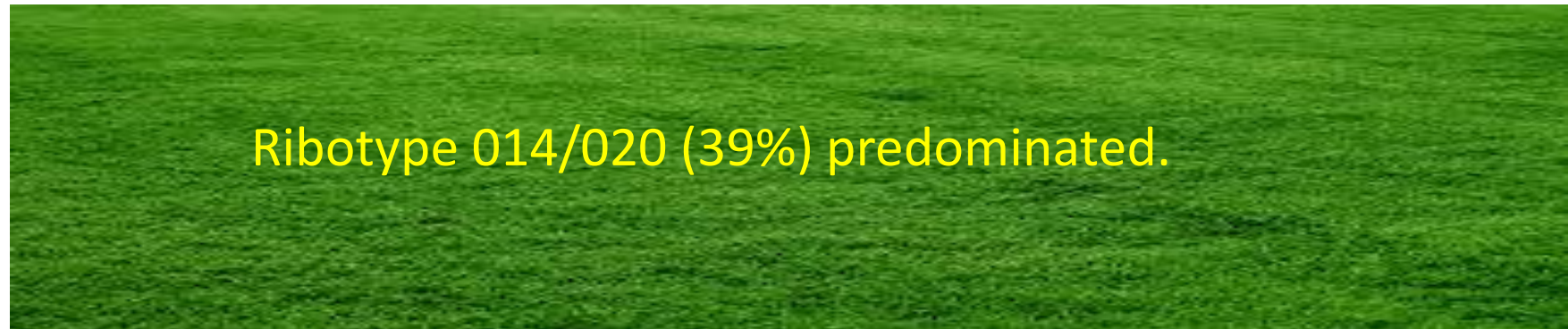


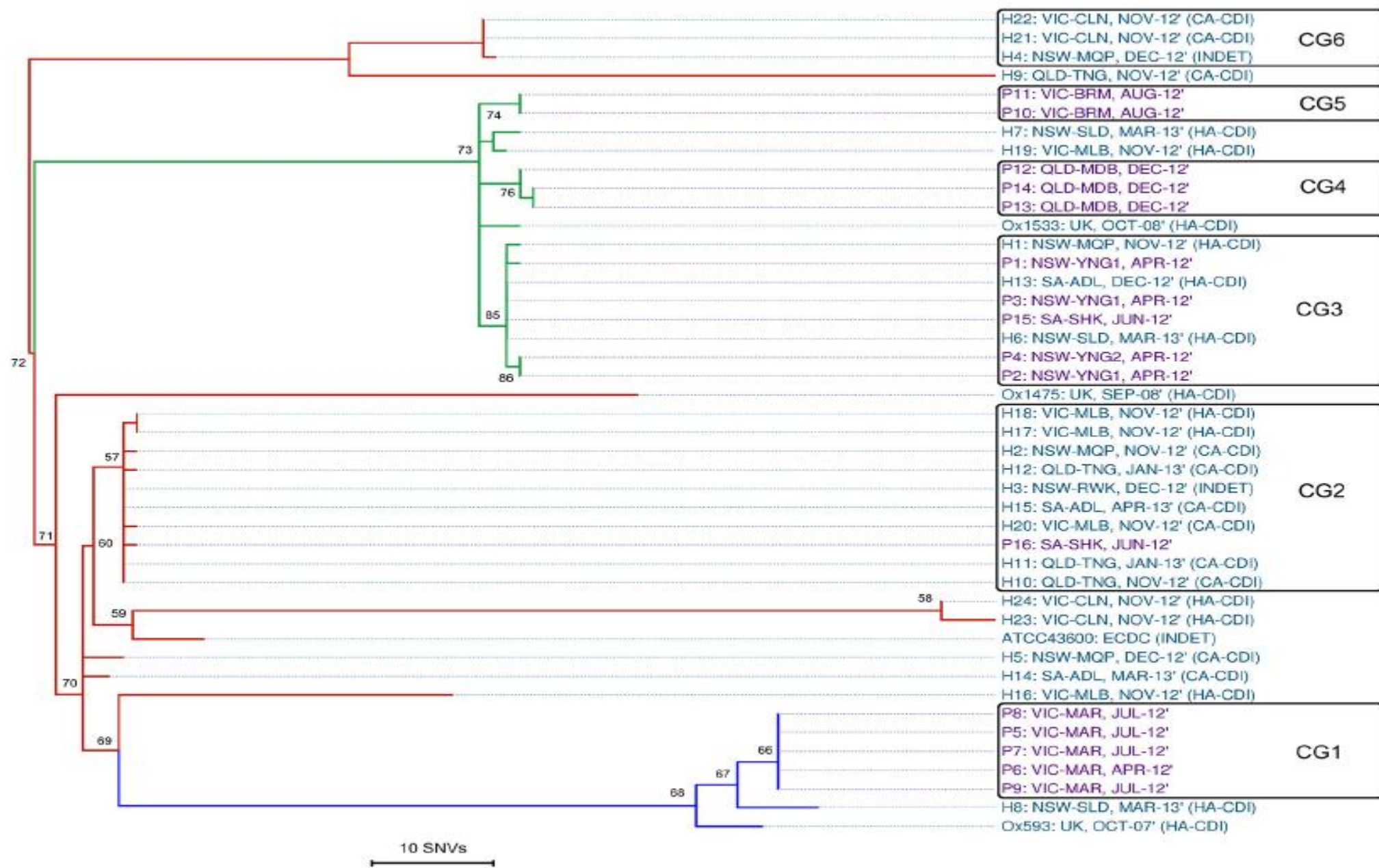
Figure 1. PCR ribotype of *C. difficile* isolates in gardening products



Variable	Variable categories	<i>C. difficile</i> number isolated (%)	Univariable model	Covariate Odds ratios (95% CI)*	
			Odds ratios (95% CI) <sup>†</sup>	Sampling site	<i>P</i> value <sup>‡</sup>
Age <sup>‡</sup>	Old lawn (n = 113)	53 (47)	Referent		
	New lawn (n = 198)	129 (65)	2.11 (1.32–3.4)	2.30 (1.16–4.57)	0.015 <sup>#</sup>
Area	Extra-large (n = 85)	53 (62)	Referent		
	Large (n = 53)	26 (49)	0.58 (0.28–1.16)	0.49 (0.16–1.49)	0.7
	Medium (n = 101)	60 (59)	0.88 (0.49–1.59)	1.02 (0.42–2.51)	0.7
	Small (n = 72)	43 (60)	0.89 (0.47–1.71)	0.88 (0.32–2.43)	0.7
Location	North (n = 161)	98 (60.9)	Referent		
	South (n = 150)	84 (56)	1.22 (0.78–1.92)	1.25 (0.61–2.59)	0.99
Season	Autumn (n = 224)	135 (60.3)	Referent		
	Winter (n = 87)	47 (54)	0.77 (0.47–1.28)	0.67 (0.28–1.62)	0.52

**Table 1.** The relationship between the prevalence of *C. difficile* in lawn and the age of the lawn, its size, sampling site, location, postcode, and season in Perth.





**FIGURE 3 |** Single nucleotide variant analysis of 44 *C. difficile* RT014.

# What are the sources/reservoirs for community-acquired CDI?

Food yes, but mainly root vegetables.

Probably more important as a vehicle rather than food *per se*.

Contamination of households – kitchens in particular.

Meat less likely an issue – adult animals OK.

Notwithstanding the above, cooking won't help!

**ANIMAL MANURE THE MAIN PROBLEM!**



# Transport of *C. difficile* into hospitals

- In patients
- In visitors
- On visitors (hands etc. and on shoe soles)
- On/in food
- Animals
- Hospital gardens and lawns

Therefore the problem remains the same:  
reduce environmental contamination.

# The way forward

- Continue to try and engage the veterinarians in One Health
- Explain to farmers/industry groups what is going on
- Involve politicians
- Look for alternative treatment agents/strategies for animals
- Restrict cephalosporin use in human medicine
- Completely ban all cephalosporins from use in production animals

**“Cephalosporins are the work of the devil, send to end the antibiotic era!”**



# Lessons from Western Australia

Many sources of *C. difficile* other than food probably equally or more important (such as lawn).

Each jurisdiction will need to look in its own back yard!

Anywhere there is animal manure there is a problem.

Not just food animals – horses\* also, maybe companion animals.

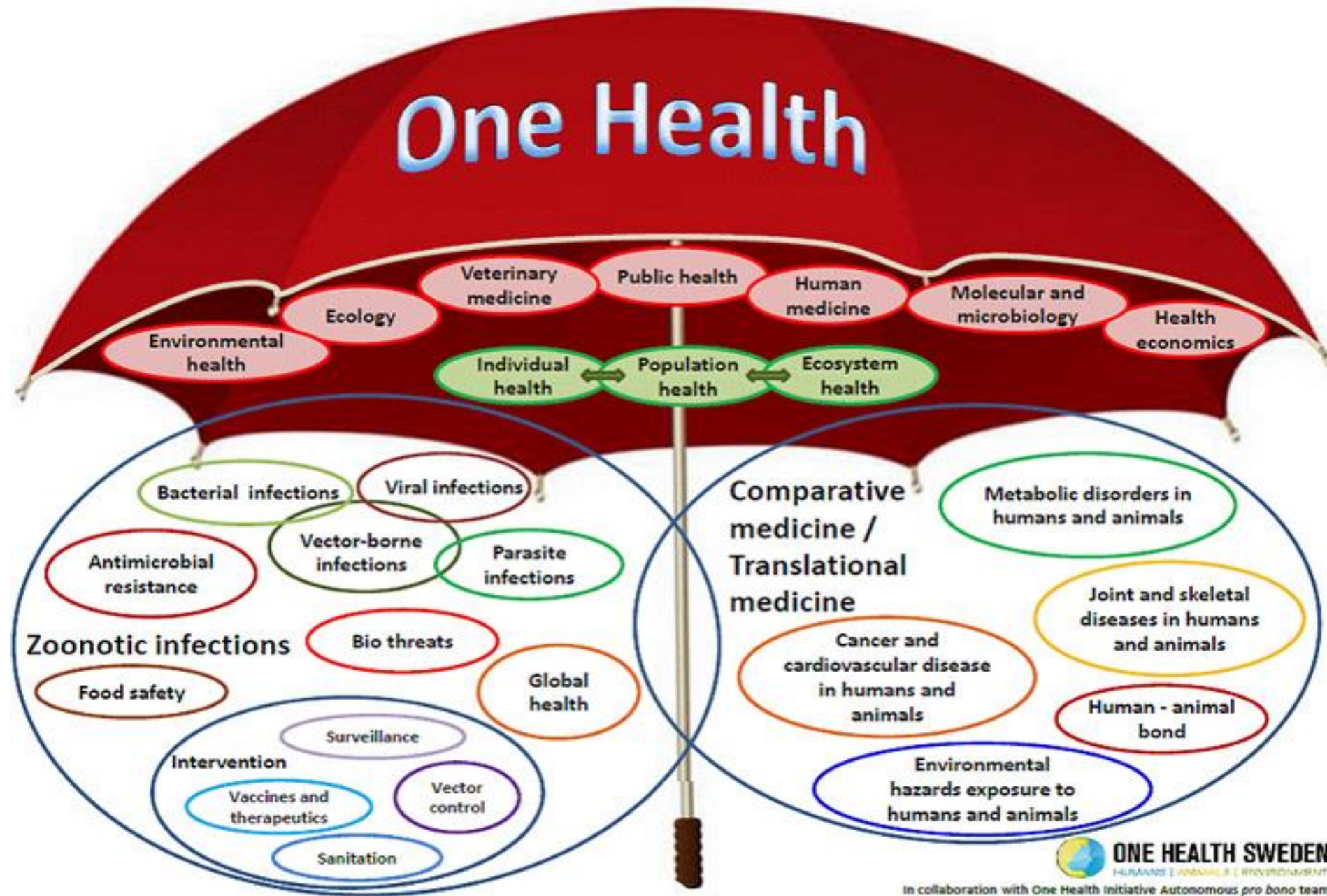
Antimicrobial use in production animals is driving this problem.

Requires a One Health approach.

\* Food in France



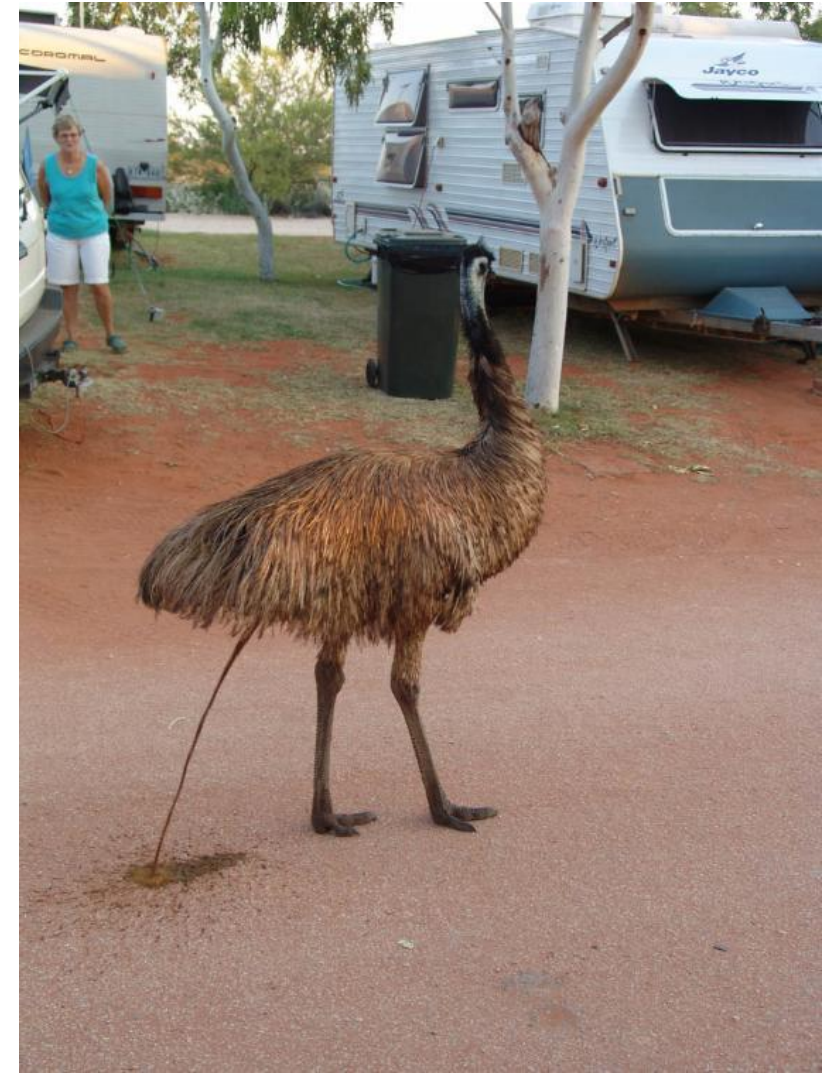
The One Health concept recognizes that the health of humans is connected to the health of animals and the environment.



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